

Review on the Production of Citric Acid through Solid-State Fermentation on Sugarcane Bagasse

Brijesh Kumar Singh¹ Deepak Kumar Patel² Ramesh Chandra Patel³ Shivangi Nigam⁴
Ashutosh Mishra⁵

^{1,2,3,4,5}Dr. Ambedkar Institute of Technology for Handicapped, Kanpur U.P., India

Abstract— For citric acid production four isolates of *Aspergillus Niger* (viz CA16,14/20,HB3 and 318) were used. Sugarcane bagasse was used as a substrate and sucrose solution used as a moistening agent. The highest citric acid production was obtained when 10 ml of 14% sucrose solution was used as moistening agent. For 11 days fermentation for all isolates of *Aspergillus Niger*.

Keywords: Citric Acid, Solid-State Fermentation, *Aspergillus Niger*, Sugarcane Bagasse

I. INTRODUCTION

Citric acid is one of the most commonly used organic acid in the food and pharmaceutical industry, which is produced mainly by submerged or surface fungal fermentation. *Aspergillus Niger* fermentation is the world's leading source for commercial production of citric acid. Solid-state fermentation is regarded as better option for citric acid production because of lower energy requirement, higher product yield with little risk of bacterial contamination, generation of less waste water and environmental concerns regarding the disposal of solid-waste. Another benefit for solid-state fermentation is that, inhibition of citric acid production due to presence of metal ions is ineffective in solid-state fermentation. Since 1920, all commercial citric acid was produced from lemon and lime juices. In 1923, at first time *Aspergillus Niger*, a fungus used for production of citric acid. Factors which affect the production of citric acid by fermentation are the nutritional composition of the media, environmental conditions, deficiency of manganese and other metals, pH, and dissolved oxygen concentration. Now a days citric acid is commercially produced by using mutant strains of *Aspergillus Niger*, and with a significant amount by *Saccharomyces lipolytic.*, large amounts of sugarcane bagasse are produced as a by-product of sugar industries. Sugarcane bagasse is the best source for solid state fermentation for production of citric acid. The study was done to investigate the possibility of using sugarcane bagasse as a substrate fortified with different quantity of sucrose solution as moistening agent and additional carbon source for solid-state fermentation for citric acid production by four isolates of *Aspergillus Niger*.

II. MATERIAL & METHOD

A. Area of study

This study was conducted at the Department of Chemical Engineering All experiments were accomplished aseptically in the laboratory of chemical engineering..

B. Collection of sample

Three samples were collected from air, bread, and soil in depth of 15 cm from Tuti islands' farms (Erika et al., 2013). Forty litres of sugarcane molasses sample were obtained from the Distillery Unit of Kennan Sugars (D.U.K.S) Company –

White Province – Sudan. The sugarcane molasses were collected in clean, durable plastic container and stored at room temperature for further uses.

C. Isolation of test microorganism (*Aspergillusniger*)

One gram of the soil sample was placed in the test tube containing 10 ml of sterile distilled water to make a soil suspension and tenfold serial dilution was made by transferring one ml of the soil suspension to another test tube containing 9 ml of sterile distilled water. This step was repeated ten times to obtain a dilution of 10⁻¹⁰. An amount of 0.1 ml from each of the first three test tubes (10⁻¹, 10⁻², and 10⁻³) was taken and placed on the plate containing Sabouraud dextrose agar medium supplemented with rose bengal to inhibit the growth of saprophytes fungi other than *A. niger*. Another plate was opened inside the laboratory of microbiology to isolate *A. niger* from the air; the third plate was inoculated with the *A. niger* from infested bread; all plates were incubated aerobically at 25°C for 72 hrs. After the incubation period, the culture characteristics were observed and the growth was examined microscopically to confirm its purity using lactophenol cotton blue stain technique (Cheesbrough, 2008).

D. Physical characteristics of the sugarcane molasses sample

The physical characteristics of sugar cane molasses, such as moisture content, ash measurement and pH, were analyzed following standard methods (APHA, 2000).

1) Moisture content and ash measurement

The moisture content and ash measurement of molasses was performed by taken 10 grams of molasses sample and oven dried in a crucible at 104°C for 30 minutes (Hubert, 2006). Then the results were calculated using the following equations:

$$\text{Moisture content(\%)} = (A - X \div A) \times 100 \quad (1)$$

$$\text{Ash (unit)} = \frac{\text{Weight of molasses before burning (A)} - \text{Weight of molasses after burning (X)}}{\text{Weight of molasses before burning (A)}} \quad (2)$$

where A is the weight of molasses before burning.

While X is the weight of molasses after.

2) The pH value

The pH value was measured before and after inoculation of molasses samples using pH meter device (pH 213 Microprocessor-based Bench pH/mV/C Meters. Hanna Instruments).

E. Production of citric acid from raw sugarcane molasses

Isolates of *A. niger* were transferred to the 15 flasks containing raw sugarcane molasses media with different concentrations, i.e., each three flasks have the equal amount of molasses (20%, 30%, 40%, 50%, and 60%) by taking 100 ml, 150 ml, 200 ml, 250 ml, and 300 ml of sugarcane molasses and the volume was completed to 500 ml using sterile distilled water. The flasks were autoclaved at 115°C for 10 minutes. An amount of 50 ml of distilled water was

added to the fungal pure culture to make a fungal suspension and then 10 ml from this suspension was transferred to the sugarcane molasses media. All flasks were incubated at 28°C for 144 hrs till 10 days. After incubation, the suspension was distilled to monitor the growth and observe the results (Elholi and Al-Delaimy, 2003).

F. Production of citric acid from sugarcane molasses with determined concentration of sucrose

In another experiment, different concentrations of sucrose (10%, 25%, 35%, and 50%) were measured using a hand refractometer device. After sterilization, an amount of 0.5 grams of urea powder was added to each flask containing sugarcane molasses with known concentration. Then 3 ml from pure *A. niger* were added to the media and the culture was incubated at 28°C for 144 hrs till 10 days. After incubation, the suspension was distilled to monitor the growth and observe the results (Dubey, 2003).

G. Detection of citric acid

The detection of citric acid was done chemically by the addition of three drops of bromocrysol green indicator to the 10 ml of distillation yield (Soccol et al., 2006).

H. Determination of citric acid concentration

Citric acid was determined by titration using 0.1N NaOH and Phenolphthalein as indicator and calculated as percentage according to the following formula (Soccol et al., 2006):

- Normality of Citric acid = normality of NaOH × NaOH volume ÷ volume of Citric acid
- Concentration of Citric acid = Citric acid normality × equivalent × 100 ÷ volume of distillation
- (Equivalent = 96, volume of distillation = 10)

III. RESULTS & DISCUSSION

A. Isolation of *Aspergillus niger*

Three isolates were isolated from three different sources air, bread, and soil using sterile culture media, the isolates were purified, examined microscopically to show its purity and characterized by its culture characteristics.

B. Physical characteristics of the sugarcane molasses sample

The physical characteristics of sugarcane molasses were determined and calculated. The present study shows that the percentage moisture content was 65%. The ash was calculated as 6.50%. While the pH shown 6.0±0.2.

These findings were in disagreement with the findings of Gasmalla et al. (2012) who reported that the pH value of obtained molasses was 5.8±0.35. The ash was 12.69% on wet weight basis. Also these findings were in disagreement with the findings of Osunkoya and Okwudinka (2011) who reported that the pH value of obtained molasses was 5.1. The ash was 8.24%.

C. Production of citric acid from raw sugarcane molasses

As can be seen in Table 1, the best yield by all strains was 37.5 ml, 37.0 ml, 35.0 ml which were obtained at concentration of 20%. At concentration 30% they were 37.0 ml, 37.5 ml, and 33.5 ml, while at concentration 40% the two strains that were isolated from soil and bread gave a similar

yield 35.0 ml and the air isolate strains gave 20.0 ml citric acid. The present study was almost in agreement with Sikander et al. (2002) who reported four *A. niger* isolates produced citric acid with the concentration of 18.86±1.8 – 42.56±2.0 g/l on 150g/l molasses sugar.

The lowest yields were obtained at the concentration of 50% as 10.0 ml, 5.5 ml, and 8.0 ml citric acid. At 60% sugarcane molasses concentration the microorganism did not exhibited any growth due to the effect of hypertonic solution. These findings were in disagreement with Sikander et al. (2002) who stated that three *A. niger* cultures gave concentrations of citric acid ranged between 58.14±2.7 – 78.18±18 g/l on 150g/l molasses sugar. Also, the present study was in agreement with Peksel and Kubicek (2003) who reported that the concentration and type of sugar influence the yield of citric acid production by *A. niger*. The present findings also were in agreement with Laboniet et al. (2010) who reported that the citric acid production increased with the increase of the fermentation period and the maximum citric acid production was found on day 13.

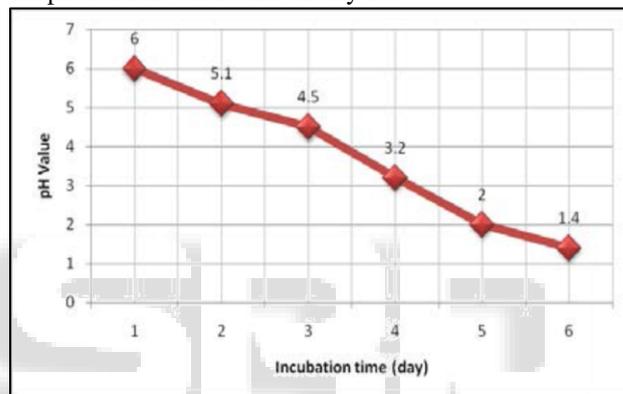


Fig. 1: Citric acid indicated by pH reduction

(2014) who stated that the production of citric acid by *A. niger*, cultured on Parkiabiglobosa fruit pulp, showed that the highest yield (1.15 g/L) of citric acid was obtained at pH 2 and it declined as the pH increased from being acidic to alkaline (pH8) with the yield of (0.86 g/L).

During the fermentation process there was a gradual reduction (Figure 1) of pH noticed in all the experiments and it indicated the production of citric acid.

	Solids/ g	Soil yield (ml)	Bread yield (ml)	Air yield (ml)
20	28.50	37.5	37.0	35.0
30	42.75	37.0	37.5	33.5
40	57.00	35.0	35.0	20.0
50	71.25	10.0	5.5	8.0
60	85.50	No production	No production	No production

Table 1. Production of citric acid from raw molasses Sugarcane Molasses concentration%

D. Production of citric acid from molasses with determined concentration of sucrose

As can be seen in Table 2, the yield of citric acid was high when using the *A. niger* which was isolated from soil at all concentrations compared with other isolates (air, bread), followed by bread isolates, then air isolates which was the lowest yield. These findings were in disagreement with Kareem et al. (2010) who stated that the inoculation of *A.*

nigeron medium supplemented with sucrose (15% w/v) gave the highest citric acid value (36.6 g/kg). Also these findings were in agreement with the same author who stated that *A. niger*, when inoculated on medium containing pineapple peels, gave 17.23 g/kg at 5 days fermentation period. The increase in citric acid production and biomass values was accompanied with a steady decrease in sugar along the incubation time.

The addition of urea as a nitrogen source did not affect the production of citric acid; this is in agreement with Sadiet al. (2011) who reported that all concentrations (0.1 to 0.6%) of ammonium sulphate, peptone and yeast extract, used as a nitrogen source, were found to be inhibitory to fungal growth, sugar utilization and citric acid production. Also, the findings of this study were in agreement with Laboniet al. (2010) who stated that in the presence of prescott salt, citric acid production was found lower than it is with the absence of prescott salt and mixed substrate prepared with molasses and pumpkin media proved to be the best and potential for citric acid production. The natural oils with high unsaturated fatty acids content when added at concentrations of 2% and 4% (v/v) to Beet Molasses (BM) medium caused a considerable increase in citric acid yield from *A. niger*. The maximum citric acid yield was achieved in surface culture in the presence of 4% olive oil after 12 days incubation.

E. Production of citric acid from molasses with determined concentration of sucrose

As can be seen in Table 2, the yield of citric acid was high when using the *A. niger* which was isolated from soil at all concentrations compared with other isolates (air, bread), followed by bread isolates, then air isolates which was the lowest yield. These findings were in disagreement with Kareem et al. (2010) who stated that the inoculation of *A. niger* on medium supplemented with sucrose (15% w/v) gave the highest citric acid value (36.6 g/kg). Also these findings were in agreement with the same author who stated that *A. niger*, when inoculated on medium containing pineapple peels, gave 17.23 g/kg at 5 days fermentation period. The increase in citric acid production and biomass values was accompanied with a steady decrease in sugar along the incubation time.

The addition of urea as a nitrogen source did not affect the production of citric acid; this is in agreement with Sadiet al. (2011) who reported that all concentrations (0.1 to 0.6%) of ammonium sulphate, peptone and yeast extract, used as a nitrogen source, were found to be inhibitory to fungal growth, sugar utilization and citric acid production. Also, the findings of this study were in agreement with Laboniet al. (2010) who stated that in the presence of prescott salt, citric acid production was found lower than it is with the absence of prescott salt and mixed substrate prepared with molasses and pumpkin media proved to be the best and potential for citric acid production. The natural oils with high unsaturated fatty acids content when added at concentrations of 2% and 4% (v/v) to Beet Molasses (BM) medium caused a considerable increase in citric acid yield from *A. niger*. The maximum citric acid yield was achieved in surface culture in the presence of 4% olive oil after 12 days incubation.

	Solids/g	Urea /g	Soil yield (ml)	Bread yield (ml)	Air yield(ml)
10	14.25	0.5	10.6	10.0	9.0
25	35.63	18.0	15.0	13.0	
35	49.88	19.5	17.0	10.5	
50	71.25	9.0	7.5	8.5	

Table 2: Production of citric acid from molasses (sucrose + urea) Sucrose%

IV. CONCLUSION

We found in this study that using sugarcane bagasse as a natural fermentation medium fortified with 10 of 15% sucrose as additional carbon source and moistening agent for citric acid production was superior when supplemented with Prescott salt. Maximum citric acid was produced by *A. niger* 318 on sugarcane bagasse in solid-state fermentation.

REFERENCES

- [1] Kubicek CP, Witteveen CFB & Visser J. 1994. Regulation of organic acid production by *Aspergilli*. In *The Genus Aspergillus* (Powell KA ed), pp. 35-145. Plenum Press, New York.
- [2] Röhr M. 1998. A century of citric acid fermentation and research. *Food Technol Biotechnol.* 36: 163-171.
- [3] Kristiansen B & Sinclair G. 1978. Production of citric acid in batch culture. *Biotechnol Bioeng.* 20: 1711-1722.
- [4] Kumar D, Jain VK, Shankar G & Srivastava A. 2003. Utilization of fruits waste for citric acid production by solid state fermentation. *Proc Biochem.* 38: 1731- 1738.
- [5] Anupama & Ravindra P. 2001. Studies on production of single cell protein by *Aspergillus niger* in solid state fermentation or rice bran. *Braze Arch Biol Technol.* 44: 79-88.
- [6] Lu M, Brook JD & Madrox IS. 1997. Citric acid production by solid state fermentation in a packed-bed reactor using *Aspergillus niger*. *Enzyme Microbe Technol.* 21: 392-397.
- [7] Pintado J, Tarra do A, Gonzalez MP & Muradov MA. 1998. Optimization of nutrient concentration for citric acid production by solid state culture of *Aspergillus niger* on polyurethane foams. *Enzyme Microbe Technol.* 23: 149-156
- [8] Rozas MG, Cordova J, Aurai R, Revah S & Favela E.
- [9] King RD & Cheetham PSJ. 1987. *Food Biotechnology*, Vol 1, pp 273-307. Elsevier Applied Science Publishers Ltd, London.
- [10] Röhr M, Kubicek CP & Kominek I. 1983. Citric acid. In 1995. Citric acid and polyols production by *Aspergillus niger* at high glucose concentration in solid state fermentation on inert support. *Biotechnol Lett.* 17(2): 214-219.
- [11] Hossain M, Brooks JD & Moddax IS. 1984. The effect of the sugar source on citric acid production by *Aspergillus niger*. *Appl Microbiol Biotechnol.* 19: 393-397.
- [12] Xu DP, Madrid CP, Röhr M & Kubcek CP. 1989. The influence of type and concentration of carbon source on production of citric acid by *Aspergillus niger*. *Appl Microbiol Biotechnol.* 30: 553-558.

- [13] Roukas T & Kotzekidou P. 1997. Pretreatment of date syrup to increase citric acid production. *Enzyme Microb Technol.* 21: 273- 276.
- [14] Bayraktar E & Mehmetoglu U. 2000. Production of citric acid using immobilized conidia of *Aspergillus niger*. *Appl Biochem Biotechnol.* 87: 117-125.
- [15] Good DW, Drouniuk R, Lawford GR & Fein JE. 1985. Isolation and characterization of a *Saccharomycopsis lipolytica* mutant showing increased production of citric acid from canola oil. *Can J Microbiol.* 31: 436-440.
- [16] Osunkoya O and Okwudinka N. 2001. Utilization of sugar refinery waste (molasses) for ethanol production using *Saccharomyces cerevisiae*. *Am J Sci Ind Res.*, 2(4): 694-706.
- [17] Pazouki M, Felse P, Sinha J and Panda T. 2000. Comparative studies on citric acid production by *Aspergillus niger* and *Candida lipolytica* using molasses and glucose. *Bioprocess Engineering*, 22(4): 353-361.
- [18] Peksel A and Kubicek C. 2003. Effect of sucrose concentration during citric acid production accumulation by *Aspergillus niger*. *Turk J Chem.*, 27:581–590.
- [19] Sikander A, Ikram U, Qadeer M and Javed I. 2002. Production of citric acid by *Aspergillus niger* using cane molasses in a stirred fermentor. *Electr J Biotechnol.*, 5 (3): 258-271.
- [20] Socoli C, Vandenberghe L, Rodrigues C and Pandey A. 2006. New perspectives for citric acid production and application. *Food Technol Biotechnol.*, 44(2): 141-165.
- [21] Thangavelu R and Murugaiyan K. 2011. An Experimental study on citric acid production by *Aspergillus niger* using *Gelidium acerosa* as a substrate. *Indian J Microbiol.*, 51(3): 289-293.